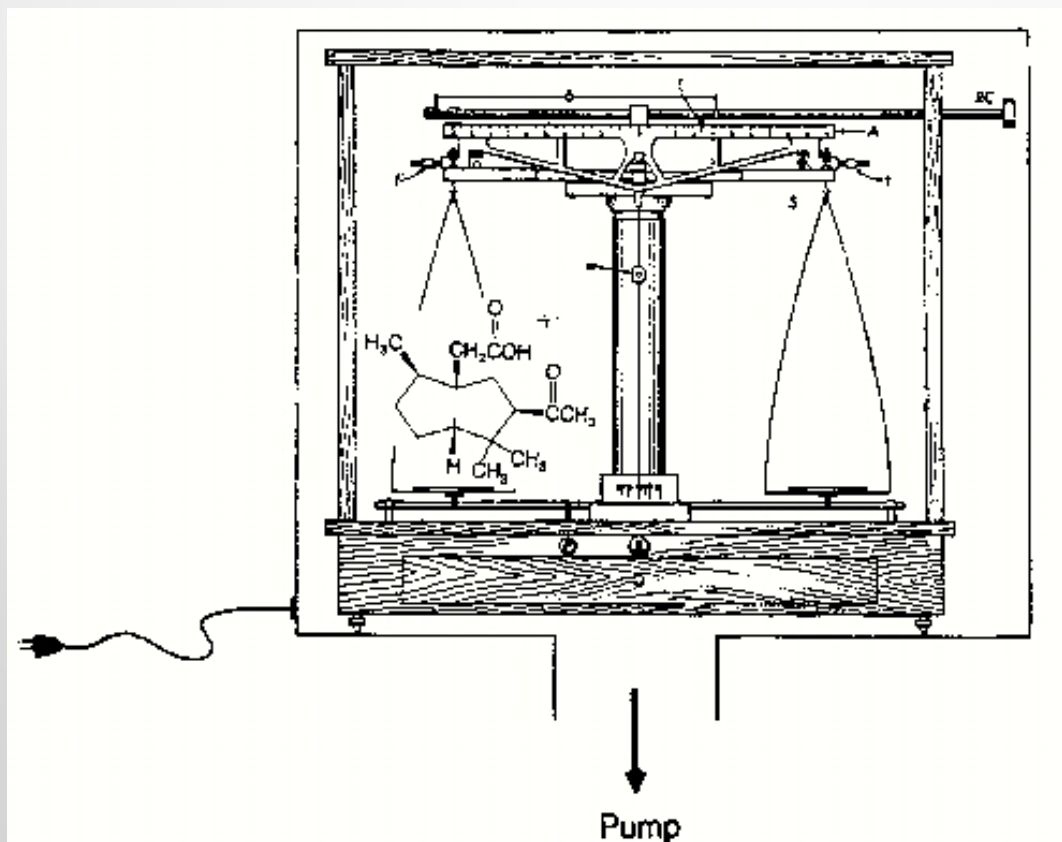


Research Opportunities in the Poutsma Group

(fun with mass spectrometers)



Mass Spectrometers



LCQ-DECA ion trap with Shimadzu UPLC



TSQ-quantum ultra (QQQ)



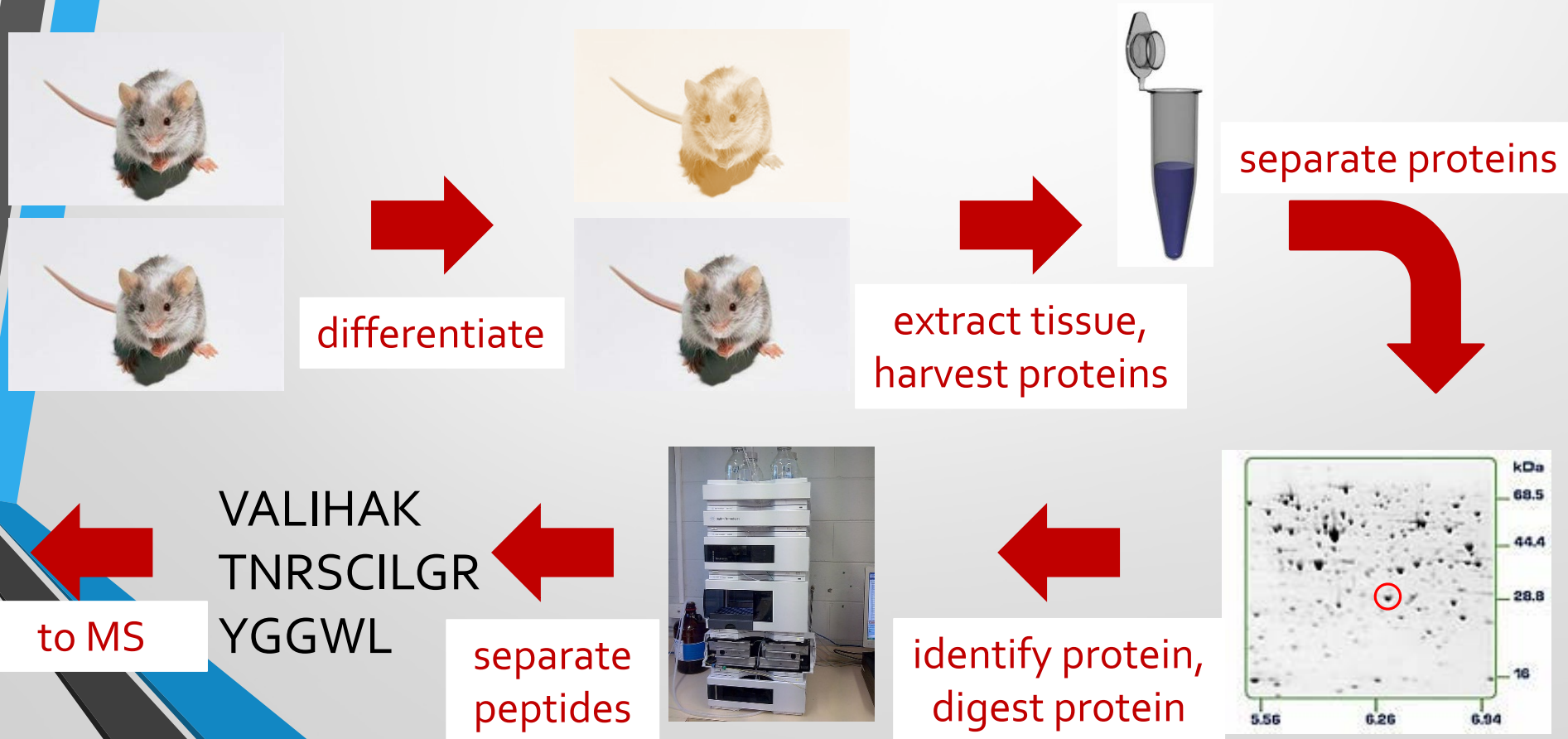
LTQ-ETD with Eksigent nano-HPLC

On-going Projects

- Gas-phase proteomics research (*using the rules*)
 - collaborative studies with Biology Dept.
 - *E. coli* infection project with Professor Williamson
- Gas-phase ion structure (*understanding/improving the rules*)
 - custom solid-phase synthesis of peptides
 - mass spectrometer fragmentation studies
 - H/D exchange of peptides and fragments
 - Infrared multiphoton dissociation (IRMPD)
- Gas-phase thermochemistry (*the basis for the rules*)
 - effects of systematic substitutions on amino acid thermochemistry
 - thermochemistry of small peptides

Proteomics

- proteomics is the study of proteins
- one might wish to determine the identity and concentration of all the proteins expressed by an organism
- or one can do a differential study of targeted proteins



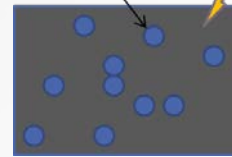
MS-based peptide sequencing

YGGWL⁺

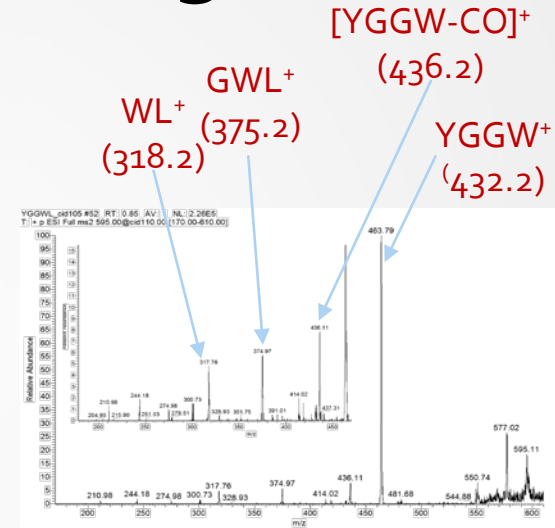


select parent
record mass
(595.6)

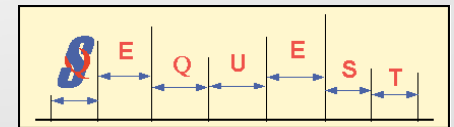
He



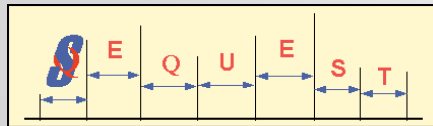
collision induced
dissociation



product ions



fingerprint
matching



protein
identification

Peak Mass (Da)	Sequence
1671.9	1-18 acetyl-GEEDVQALVVDNGSGNVK
928.45	19-28 AGVAGDDAPR
1186.67	29-39 SVFPSIVGRPK
2522.16	40-62 NPGIMVGMEEKDAFVGDEAQTQR
800.53	62-68 RGILTK
1946.91	69-84 YPIEHGIVTNWDDMEK
1515.73	85-95 IWHITFYNELR
1926.03	96-113 AAPEEHPVLLTEAPLNPK
3169.49	148-177 TTGIVLDSGDGVSHTVPIYEGYALPHAIR
644.4	178-183 LDLAGR
1012.5	184-191 DLTEYLMK
595.7	192-196 YGGWL
1046.51	197-206 GYGSTSAEK
1129.65	207-215 ENRDIKEK
1645.72	216-228 LCYALNFDDEMK
1776.86	239-254 SYELPDGNIITVGNER
1796.92	255-269 FRCPEALFQPSFLGK
1617.83	270-284 EAAGIHTTTFSIKK
1033.56	284-291 KCDVDIRK
2245.08	292-312 DLYGNIVLSGGTTMYEGIGER
1177.61	316-326 DITTLAPSTMK
1136.72	327-336 IKVWPPERK
1517.68	360-372 EEYDESGSIVHR

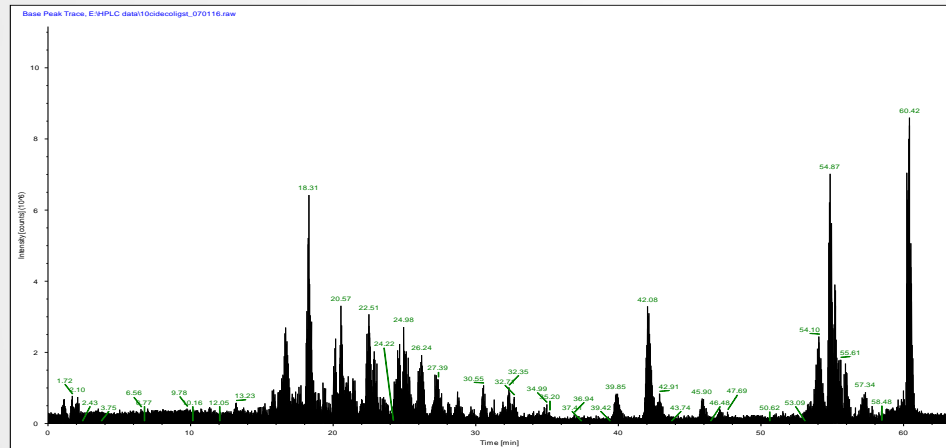
peptide
identification

Actual real-life samples

- *M. Smegmatis* infection study
 - Many novel bacteriophage viruses can infect the soil bacterium *M. smegmatis*.
 - *M. Smegmatis* is a non-pathogenic model system for *M. tuberculosis*
 - we freeze *M. Smegmatis* cells at varying time points after infection by novel phages that were discovered in the PhageLab Freshman Biology Lab experience.
 - We lyse the cells, harvest the proteins, and digest with trypsin.
 - We then perform shotgun proteomics experiments aim to identify all of the proteins being expressed by the bacterium and the virus at varying time points.
 - By determining which viral proteins are expressed at different times after infection, we can begin to classify sequenced proteins of unknown function as regulatory (early time points) or structural (late time points)
 - In addition to the information gleaned from the viral proteins, we can also begin to look for bacterial responses to infection.

Preliminary Results

E. coli sample: 15 minutes after infection with T7 phage
>3800 proteins, >20,000 peptides

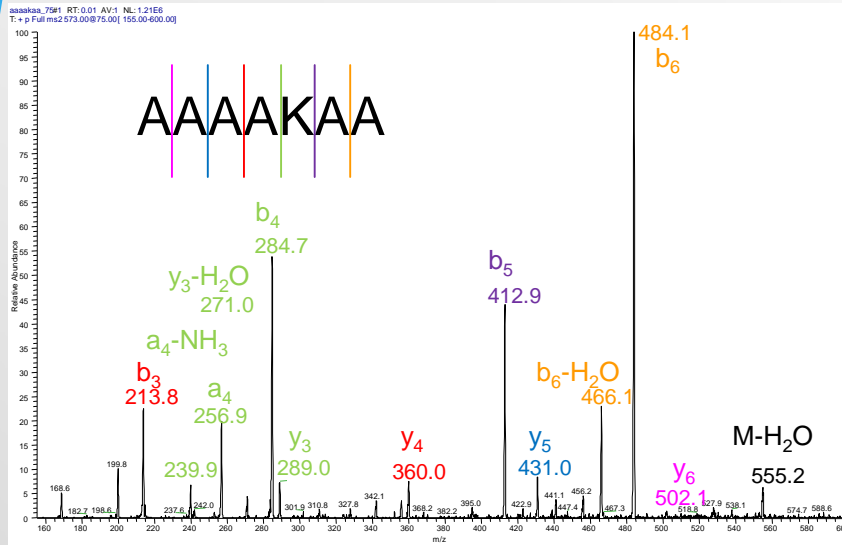


time	# proteins	representative proteins
15 min	21	DNA primase/helicase, DNA-directed DNA polymerase peptidoglycan hydrolase gp 16
30 min	27	endonuclease I portar protein
45 min	30	bacterial RNS polymerase inhibitor exonuclease inhibitor of dGTPase terminase (large and small subunits)
60 min	26	capsid assembly scaffolding protein nucleotide kinase tail tubular protein

On-going Projects

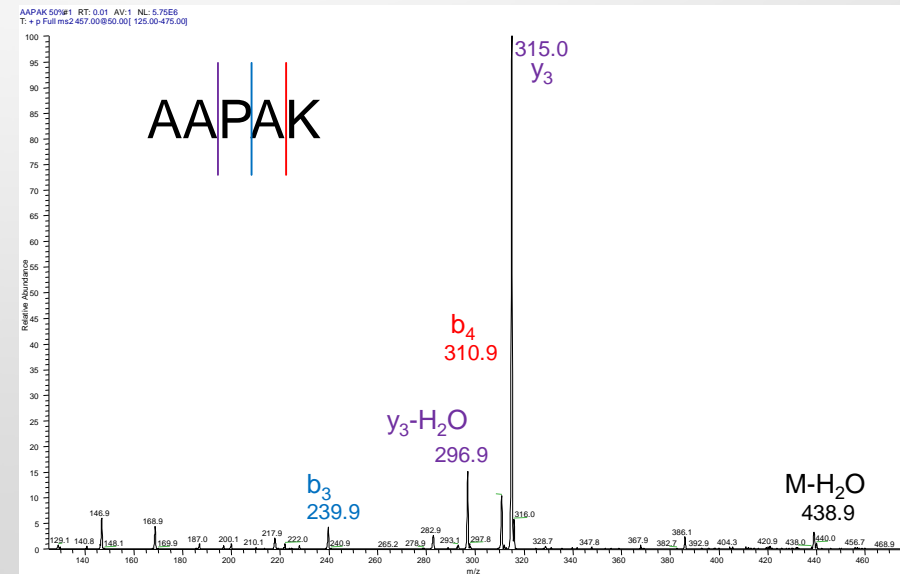
- Gas-phase proteomics research (using the rules)
 - collaborative studies with Biology Dept.
 - *H. pylori* project with Professor Forsyth
 - *E. coli* infection project with Professor Williamson
- Gas-phase ion structure (understanding/improving the rules)
 - custom solid-phase synthesis of peptides
 - mass spectrometer fragmentation studies
 - H/D exchange of peptides and fragments
 - Infrared multiphoton dissociation (IRMPD)
- Gas-phase thermochemistry (the basis for the rules)
 - effects of systematic substitutions on amino acid thermochemistry
 - thermochemistry of small peptides

Selective Cleavages (improving the rules)



peptides normally fragment randomly
along peptide backbone

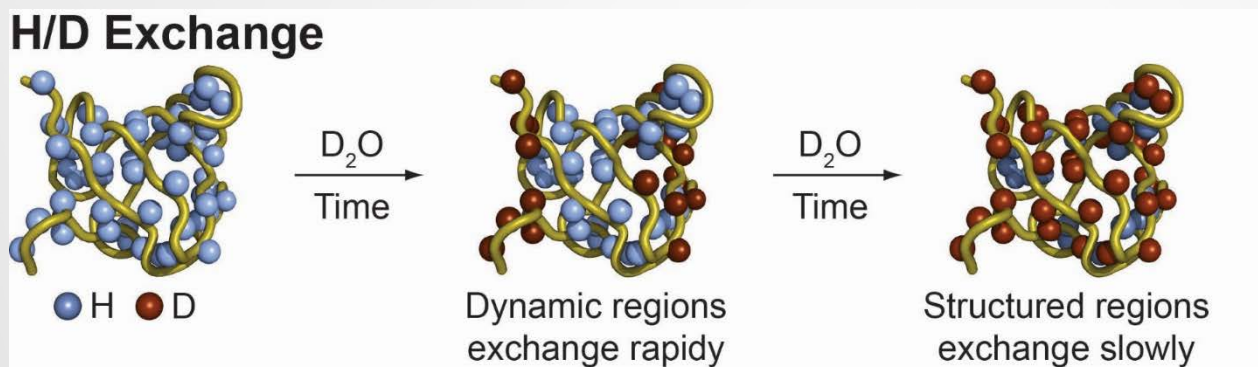
some residues such as proline produce
selective cleavages that can
confound searching algorithms



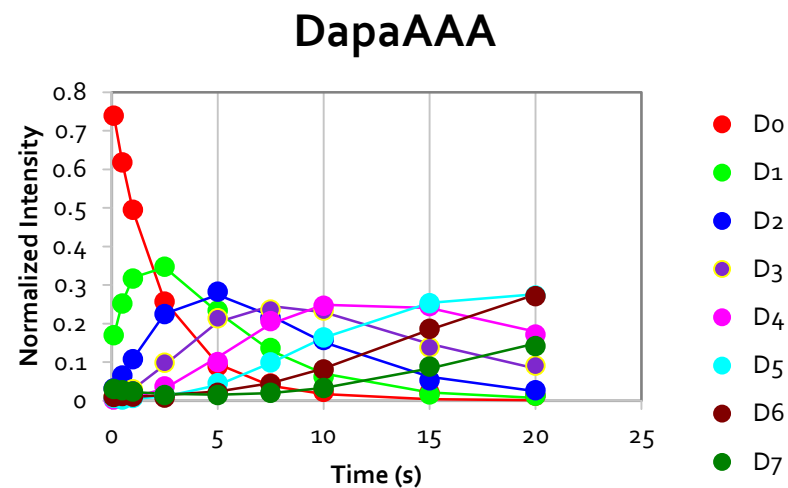
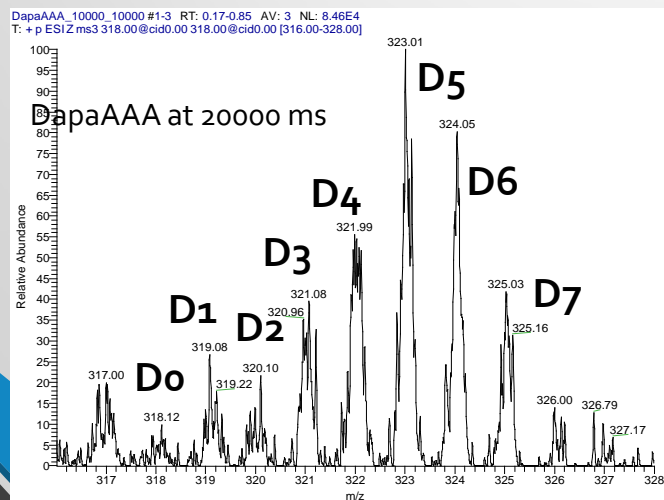
Hydrogen-deuterium Exchange

(understanding the rules)

the rate at which a gas-phase ion substitutes H for D gives an indication of the availability of exchangeable hydrogens, and thus an indirect indication of structure



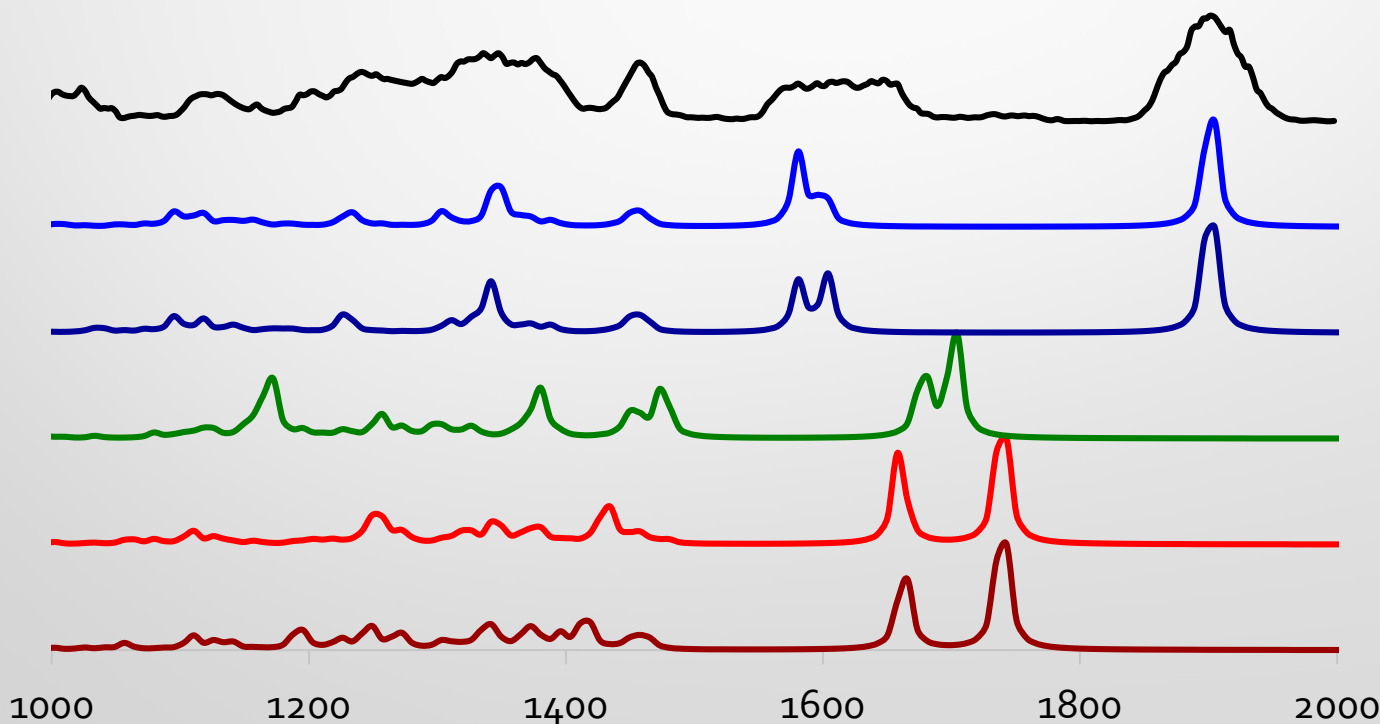
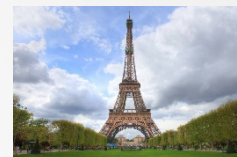
we can measure rates for H/D exchange in our ion trap mass spectrometer



IRMPD: Vibrational Spectroscopy

(understanding the rules)

- CLIO (Orsay) and FELIX (Nijmegen) FEL's
- infrared action spectroscopy of ions
- gives direct information of structure

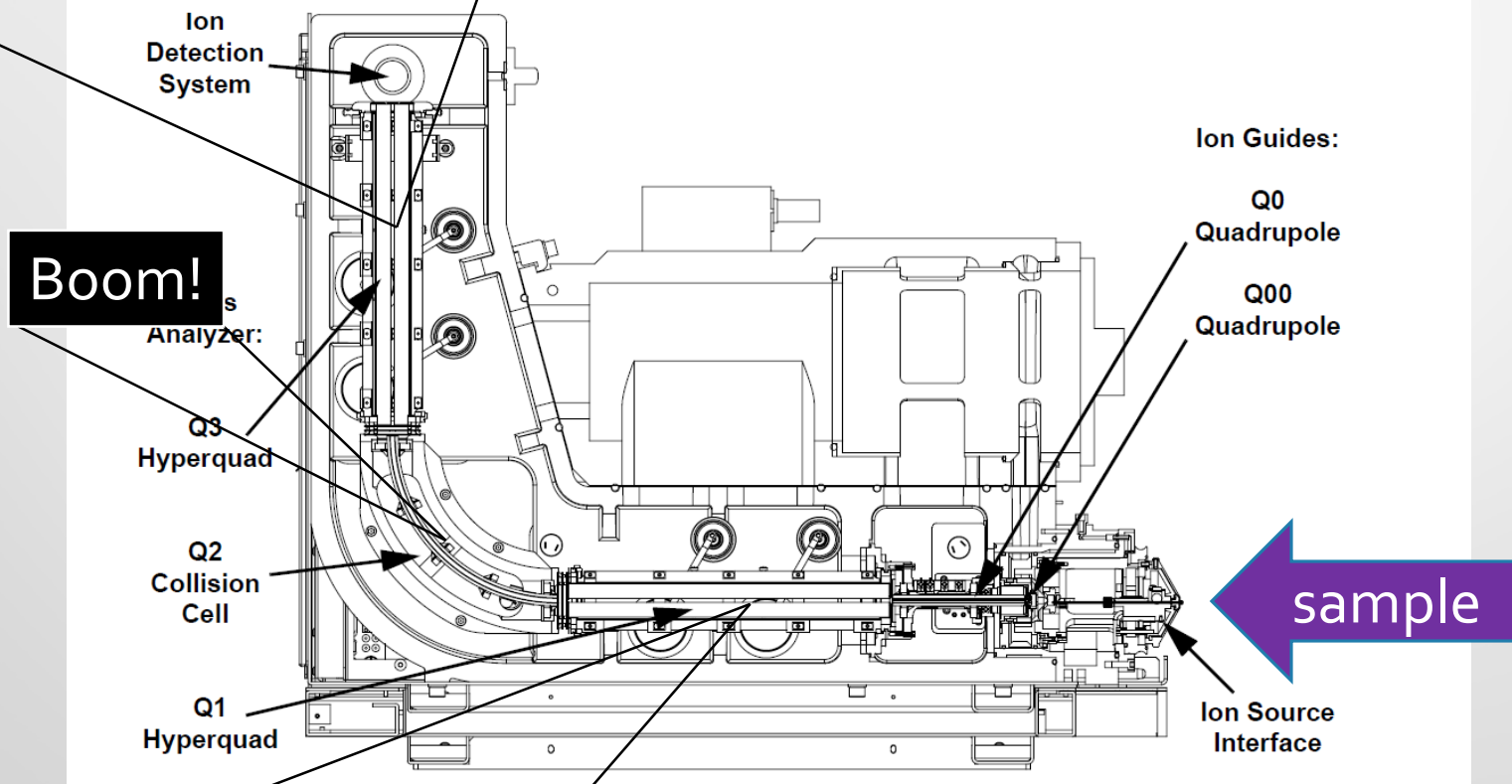


On-going Projects

- Gas-phase proteomics research (using the rules)
 - collaborative studies with Biology Dept.
 - *H. pylori* project with Professor Forsyth
 - *E. coli* infection project with Professor Williamson
- Gas-phase ion structure (understanding/improving the rules)
 - custom solid-phase synthesis of peptides
 - mass spectrometer fragmentation studies
 - H/D exchange of peptides and fragments
 - Infrared multiphoton dissociation (IRMPD)
- Gas-phase thermochemistry (the basis for the rules)
 - effects of systematic substitutions on amino acid thermochemistry
 - thermochemistry of small peptides

ESI-QQQ instrument kinetic method studies (the basis for the rules)

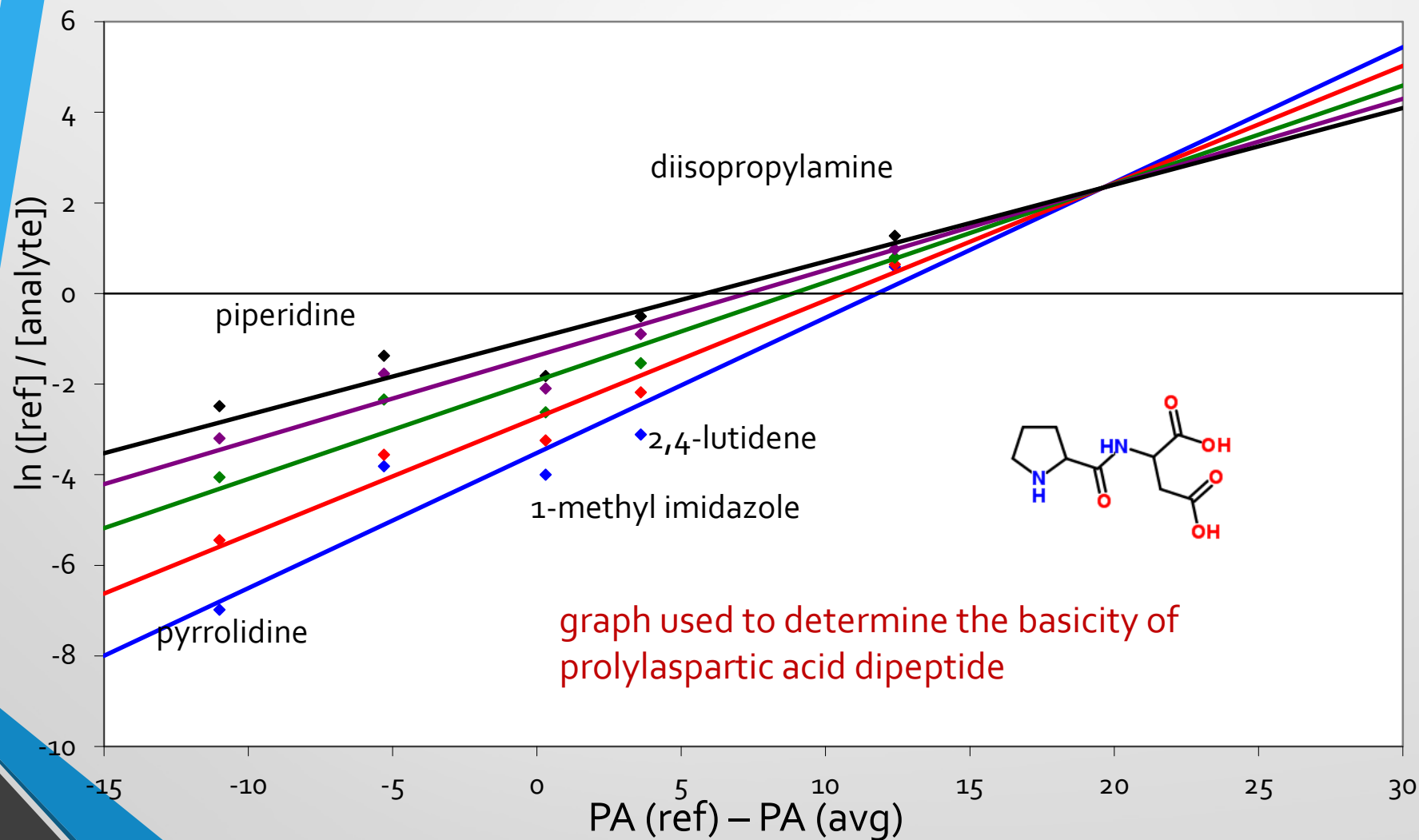
[peptide-H⁺ or base-H⁺]



[peptide₁----H⁺---base₂]⁺

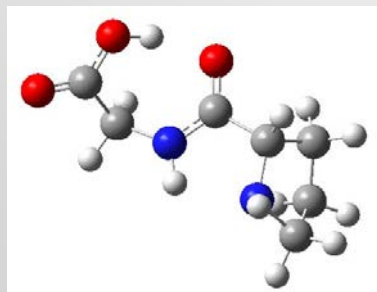
measuring the fragment intensities of competitive cleavage of proton-bound dimer ions gives us acid-base properties for peptides

Pro-Asp: kinetic method plot

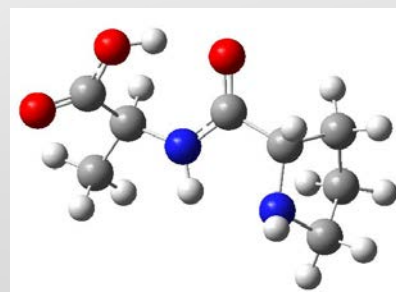


Computational Studies

- We use computational chemistry to support and guide our experimental work.
- Collaborator: Prof. Jennifer Poutsma (ODU)



Pro-Gly



Pro-Ala